# Molecular Analysis of the MUT gene in Filipino Patients with Methylmalonic Acidemia

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#### ABSTRACT

Introduction. Methylmalonic acidemia (MMA) is an autosomal recessive inborn error of metabolism resulting from defects in the nuclear encoded mitochondrial enzyme methylmalonyl-CoA mutase. This study characterizes for the first time the genotype of Filipino patients with MMA.

Methods. Clinical data were collected from 3 patients diagnosed with MMA at the Department of Pediatrics of the Philippine General Hospital from January 2002 to June 2008. The diagnosis was confirmed by urine organic acid analysis using gas chromatography – mass spectrometry (GC-MS). Molecular analysis of the MUT gene was subsequently performed using DNA from dried blood spots or peripheral blood of patients, PCR amplification and direct sequence analysis.

Results. The patients presented classically with progressive encephalopathy, metabolic acidosis and secondary hyperammonemia in the early neonatal period. Urine amino and organic acid screens showed increased glycine, methylmalonic acid and other secondary metabolites for methylmalonic aciduria. Mutations detected in the MUT gene analysis [c.1595G>A (p.R532H), c.2011G>A (p.V671I), c.322C>T (p.R108C), c.982C>T (p.L328F) and c.1280G>A (p.G427D)] were compound heterozygous in all patients.

Conclusion. Our results show the genetic heterogeneity in Filipino MMA patients and helped emphasize the importance of molecular diagnosis particularly in the genetic counseling of the patients and their families.

Key Words: Methylmalonic Acidemia, Metabolic Disorders, Methylmalonyl-CoA mutase, MMA, MUT

#### Introduction

Methylmalonic acidemia (MMA; OMIM 251000) is an autosomal recessive disorder caused by inadequate function of the methylmalonyl-CoA mutase, a mitochondrial enzyme involved in the metabolism of certain amino acids,

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**Figure 1.** Simplified pathway illustrating the metabolic blockage points of MMA. The mut<sup>0</sup> form of MMA results in a complete blockage of methylmalonyl-CoA mutase producing a toxic build-up of acyl-CoA precursors and their corresponding acylcarnitines, propionyl-carnitine, and methylmalonyl-carnitine.<sup>1</sup>

Methylmalonyl-CoA mutase is encoded by the MUT gene (GenBank NG\_007100) which maps to chromosome 6p12-21.2.<sup>4</sup> MUT has 13 exons spanning over 35 kb.<sup>5</sup> The open reading frame consists of 2.7 kb, encoding 750 amino acids and has a 32-amino-acid N-terminal mitochondrial leader sequence that forms the mitochondrial targeting sequence. To date, about 200 disease-causing mutations in the human MUT gene have been reported.<sup>6-8</sup>

The estimated worldwide incidence of MMA is between 1:50,000 to 1:100,000.<sup>9-14</sup> In countries offering expanded neonatal metabolic screening, cases can be detected using tandem mass spectrometry. In the Philippines, there is routine testing for 5 disorders such as congenital hypothyroidism, congenital adrenal hyperplasia, phenylketonuria, galactosemia and G6PD deficiency. Expanded neonatal metabolic screening is not yet available locally. Eight (8) patients have, however, been clinically diagnosed. Seven of these eight patients died secondary to infection and

intractable metabolic acidosis. The only surviving patient is presently being closely followed up at the Metabolic Clinic of the Department of Pediatrics, Philippine General Hospital (PGH).

This study describes the genotype of three Filipino MMA patients drawing attention to the importance of molecular diagnosis.

Cases

## Methods

Three unrelated patients of Filipino descent clinically diagnosed with MMA by the Section of Genetics, Department of Pediatrics, PGH from January 2002 to June 2008 were enrolled in this present investigation. There were no consanguineous marriages among the parents of these families. All families provided informed consent for the study.

Biochemical data clinched the diagnosis of MMA for these patients. Urine amino acid screen using High Voltage Electrophoresis profiles showed increased glycine and increased methymalonic acid on fast blue-B staining. Urine organic acid analysis further supported the diagnosis of MMA with increased levels of methylmalonic acid and other secondary metabolites such as methylcitrate.

## Molecular Analysis

Genomic DNA was extracted from dried blood spots or peripheral blood, in accordance with standard protocols, using the QIAamp Blood DNA Mini Kit (QIAGEN Inc.,

Table 1. Clinical features of th	e three MMA patients
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Valencia, Calif.).

The primers used to sequence the MUT gene were as described with minor modifications to the reaction conditions.<sup>14</sup> This allowed the analysis of the whole coding sequence including intron-exon borders and untranslated regions. PCR products were bi-directionally sequenced using the big dyeTM terminator cycle sequencing ready reaction kit and the products were subsequently separated on an ABI PRISM 3730xl electrophoresis system (Applied Biosystems, USA).

### **Results and Discussion**

Clinical data was available for all patients. Common clinical and biochemical features showed methylmalonic aciduria, acute episodes of severe metabolic acidosis, lethargy, poor suck/cry and activity, vomiting, encephalopathy, anemia, leukopenia and hyperammonemia. All clinical presentations appeared at 2 days of age and were initially managed as infection. Despite marked improvement with medical management, two of the patients included in this study died after 2 months to one year of age due to sepsis secondary to severe pneumonia and intractable metabolic acidosis. The only surviving patient is presently being closely followed up and given a low protein diet, carnitine supplementation, and metronidazole to reduce methymalonic acid production from gut flora. This patient is also being given supportive management during periods of disease (Table 1). Clinical data on age at onset and diagnosis, clinical findings, management and outcome were

Patient	Body Weight (kg)	Age of Gestation (weeks)	Age at onset/ diagnosis	Clinical Course
MMA-1 (5mo/M)	2.5	28	2d / 18d	<ul> <li>(+)FHx: 2 spontaneous abortions; 2nd liveborn died at 10d – irritability, vomiting, hypotonia – unknown diagnosis</li> <li>At birth: poor cry/activity, hypotonia, vomiting, leukopenia, thrombocytopenia, uncompensated metabolic acidosis.</li> <li>At 2 mos: alert, active, good tone, development at par with age.</li> <li>Died at 5mos: sepsis due to severe pneumonia, intractable metabolic acidosis after 48 hours.</li> </ul>
MMA-2 (3yo/M)	2.59	38	2d / 11d	At 11d: vomiting, poor suck/cry, progressive encephalopathy, severe metabolic acidosis, hyperammonemia, ketonuria, anemia, leukopenia, hypotonia At 2 mos: infectious diarrhea, severe metabolic acidosis At 4 mos: seizures, CT scan – corpus callosum dysgenesis, developmental delay. Died at 3 yrs: sepsis due to severe pneumonia, severe metabolic acidosis after 72 hrs
MMA-3 (4yo/F)	2.7	38	2d / 1mo	At 2 d: poor suck/ activity, severe metabolic acidosis At 1 mo: difficulty of breathing At 3 yrs: sepsis due to gastroenteritis, ketonuria, hyperammonemia, severe metabolic acidosis At present: development at par, carnitine, vit B12, protein restriction diet, metronidazole, regular monitoring

further analyzed and have been detailed elsewhere.<sup>15</sup>

To elucidate the molecular background of MMA in the 3 unrelated patients, sequence analyses of the MUT gene was done. The identified mutations were c.1595G>A, c.2011A>G, c.322C>T, c.982C>T and c.1280G>A. All the five mutations identified were compound heterozygous in all patients (Figures 2-3). Three (c.322C>T, c.982C>T and c.1280G>A) of the five identified mutations were previously reported to be disease-causing alleles (Table 2A).<sup>8,16,17</sup>

Table 2A. List of pathogenic mutations identified

	Mutation	Consequence	Reference
Patient 1	?	?	?
Patient 2	c.322C>T	p.R108C	Worgan et al 2006 <sup>17</sup>
	c.982C>T	p.L328F	Acquaviva et al 200518
Patient 3	c.1280G>A	p.G427D	Worgan et al 2006 <sup>17</sup>
	?	?	?

Table 2B. List of polymorphisms identified

	SNP	Allele(s) Present	AA Change	Reference(s)
Patient 1	c.1595G>A	Hetero G/A	Nonconservative (p.R532H)	Crane & Ledley 1992 <sup>16</sup> ; Worgan et al 2006 <sup>17</sup>
	c.2011G>A	Hetero A/G	Conservative (p.V671I)	Crane & Ledley $1992^{16}$ ; Worgan et al $2006^{17}$
Patient 2	?	?	?	?
Patient 3	c.1595G>A	Hetero G/A	Nonconservative (p.R532H)	Crane & Ledley $1992^{16}$ ; Worgan et al $2006^{17}$

The c.322C>T nucleotide substitution was identified in patient 2, causing an arginine to cysteine change at amino acid 108 (p.R108C) in one allele of the MUT gene. This mutation has been identified in 60% of Hispanic patients which is not surprising since Philippine history tells us of 3 centuries of Spanish colonization (1521-1898) with European missionaries and immigrants steadily flowing to the colony during the Spanish conquest in the 16th century.<sup>17,18,19</sup>



**Figure 2.** Chromatograms showing A. the previously reported polymorphisms c.1595G>A responsible for p.R532H and B. c.2011G>A responsible for p.V671I; and C. the previously reported missense mutation c.1280G>A responsible for p.G427D (GenBank M65131.1).

The c.982C>T change producing a leucine to phenylalanine change at amino acid 328 (p.L328F) was also found in patient 2 (Figure 3). This mutation was previously reported in Europeans and is said to alter the folding or the structural stability of the protein.<sup>8</sup>



**Figure 3.** Direct sequencing results of MMA Patient 2 showing the pathogenic mutations A. c.322C>T responsible for p.R108C; and B. c.982C>T responsible for p.L328F (GenBank M65131.1).

In 2006, Worgan et al. identified only in Asian patients, the missense mutation c.1280G>A, changing glycine to aspartic acid at amino acid 427 (p.G427D), to affect a highly conserved amino acid in the linker region that connects the substrate-binding N-terminal  $\beta/\alpha$  barrel domain and the cofactor (adenosylcobalamin) binding C-terminal ( $\alpha$ , $\beta$ ) domain. Patient 3 was identified to be heterozygous for this mutation (Figure 2). This finding is interesting since this suggests that Filipinos may have a similar mutation spectrum as our Asian neighbors.

The c.1595G>A transition changing the amino acid arginine to histidine at position 532 (p.R532H) of the MUT gene was found in patients 1 and 3. The p.R532H, however, is a non-conservative substitution that does not interfere with enzyme activity suggesting that it occurs in a region of the protein not intimately involved with function (Figure 2).<sup>16,20</sup> Patient 1 also presented with a c.2011G>A change causing a valine to isoleucine change at amino acid 671 of the MUT gene (p.V671I). This mutation was found to be conservative by Crane et al in 1992. Both these mutations, c.1595G>A and c.2011G>A, have been found to be common polymorphisms identified in the coding exons of the MUT gene (Table 2B).<sup>17</sup>

The study, therefore, was only able to identify both disease causing mutations in patient 2 and only one in patient 3. It is likely that the other mutations not identified in patients 1 and 3 are present in non-coding or regulatory regions of the MUT gene or in genes required for the provision of cofactor B12 since these areas were not analyzed.<sup>21</sup>

## Conclusion

The frequency of metabolic disorders in the Philippines is unknown with these diseases often presenting as a diagnostic challenge for pediatricians. Therefore, a substantial proportion of cases remain undiagnosed or misdiagnosed with serious consequences such as disability or death even before a diagnosis is made. Since genetic heterogeneity is high for MMA with several disease causing alleles repeatedly reported in different populations, direct DNA analysis therefore, allows directed mutational analyses for our own population.<sup>17</sup>

Although there is no genotype-phenotype correlation, this study still helps show the genetic heterogeneity for our patients with MMA. The molecular findings allow proper genetic counseling for these patients and highlight the potential for early prenatal diagnosis for at risk families.

#### Acknowledgments

The authors gratefully acknowledge the patients with MMA and their families for their cooperation and willingness to participate. The authors thank the reviewers for their constructive and detailed comments. We also express sincere gratitude to Kahlil dela Cruz-Rama, Michelle Demata-Rana, Aster Lynn Sur, Drs. Conchita Abarquez and Karl de Dios for their technical assistance and Mead Johnson Nutritionals for providing the nutritional support of our patients. This work was supported by the Institute of Human Genetics, National Institutes of Health Philippines.

#### References

- 1. Wikoff WR, Gangoiti JA, Barshop BA, Siuzdak G. Metabolomics Identifies perturbations in human disorders of proprionate metabolism. Clin Chem 2007;53:2169-2176.
- Willard HF, Rosenberg LE. Inherited deficiencies of human methylmalonyl-CoA mutase activity: reduced affinity of mutant apoenzyme for adenosylcobalamin. Biochem Biophys Res Commun. 1977;78:927-934.
- Hori D, Hasegawa Y, Kimura M, Yang Y, Verma IC, Yamguchi S. Clinical onset and prognosis of Asian children with organic acidemias, as detected by urinary organic acids using GC/MS, instead of mass screening. Brain Dev. 2005;27:39-45.
- Ledley FD, Lumetta MR, Zoghbi HY, Van Tuinen P, Ledbetter SA, Ledbetter DH. Mapping of human methylmalonyl-CoA mutase (MUT) locus on chromosome 6. Am J Hum Genet. 1988;42:839-846.
- 5. Nham SU, Wilkemeyer MF, Ledley FD. Structure of the human methylmalonyl-CoA mutase (MUT) locus. Genomics. 1990;8:710-716.
- Jung JW, Hwang IT, Park JE, et al. Mutation analysis of the MCM gene in Korean patients with MMA. Mol Genet Metab. 2005;84:367-370.
- Martinez MA, Rincon A, Desviat LR, Merinero B, Ugarte M, Perez B. Genetic analyses of three genes causing isolated methylmalonyl acidemia: identification of 21 novel allelic variants. Mol Genet Metab. 2005;84:317-325.
- 8. Acquaviva C, Benoist JF, Pereira S, et al. Molecular basis of methylmalonyl-CoA mutase apoenzyme defect in 40 European patients affected by mut(o) and mut-forms of methylmalonic acidemia: identification of 29 novel mutations in the MUT gene. Hum Mutat. 2005;25(2):167-76.
- Coulombe JT, Shih VE, Levy HL. Massachusetts Metabolic Disorders Screening Program. II. Methylmalonic aciduria. Pediatrics. 1981;67:26– 31.
- Lemieux B, Auray-Blais C, Giguere R, Shapcott D, Scriver CR. Newborn urine screening experience with over one million infants in the Quebec Network of Genetic Medicine. J Inherit Metab Dis. 1988;11:45–55.
- 11. Sniderman LC, Lambert M, Giguere R, et al. Outcome of individuals with low-moderate methylmalonic aciduria detected through a neonatal screening program. J Pediatr. 1999;134:675–80.
- 12. Shigematsu Y, Hirano S, Hata I, et al. Newborn mass screening and

selective screening using electrospray tandem mass spectrometry in Japan. J Chromatogr B Analyt Technol Biomed Life Sci. 2002;776:39–48.

- Chace DH, DiPerna JC, Kalas TA, Johnson RW, Naylor EW. Rapid diagnosis of methylmalonic and propionic acidemias: quantitative tandem mass spectrometric analysis of propionylcarnitine in filterpaper blood specimens obtained from newborns. Clin Chem. 2001;47: 2040–4.
- 14. Sakamoto O, Ohura T, Matsubara Y, Takayanagi M, Tsuchiya S. Mutation and haplotype analyses of the MUT gene in Japanese patients with methylmalonic acidemia. J Hum Genet. 2007;52:48-55.
- Chiong MA, dela Cruz-Rama K, Demata MA, et al. Methylmalonic acidemia in 2 Filipino children. Acta Medica Philippina 2008;42(2):61-65.
- 16. Crane AM, Jansen R, Andrews ER, Ledley FD. Cloning and expression of a mutant methylmalonyl coenzyme A mutase with altered cobalamin affinity that causes mut methylamlonic aciduria. J Clin Invest. 1992;89:385-391.
- 17. Worgan LC, Niles K, Tirone JC, et al. Spectrum of Mutations in mut Methylmalonic Acidemia and Identification of a Common Hispanic Mutation and Haplotype. Hum Mutat. 2006;27(1):31-43.
- Acquaviva C, Benoist JF, Callebaut I, et al. N219Y, a new frequent mutation among mut<sup>o</sup> forms of methylamlonic acidemia in Caucasian patients. Eur J Hum Genet. 2001;9:577-582.
- Agoncillo TC. History of the Filipino People, 8th ed. Quezon City: Garotech Publishing. 1990.
- Crane AM, Ledley FD. Clustering of mutations in methylmalonyl-CoA associated with mut methylmalonic acidemia. Am J Hum Genet. 1994;55:42-50.
- 21. Wilkemeyer M, Crane A, Ledley F. Differential diagnosis of mut and cbl Methylmalonic aciduria by DNA-mediated gene transfer in primary fibroblasts. J Clin Invest. 1991;87:915-918.